

Cyclostome Carbonic Anhydrase. Purification and Some Properties of the Enzyme from Erythrocytes of Lamprey*

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Since carbonic anhydrase is widely distributed in nature¹ comparisons of the enzyme from different species can give valuable information regarding various evolutionary aspects. The mammalian erythrocyte carbonic anhydrases have approximately the same molecular weights, about 29 000–30 000. This is also the case for the studied submammalian erythrocyte carbonic anhydrases,² except the elasmobranch carbonic anhydrases which have molecular weights of 36 000–39 000.³

In order to see whether the heavier elasmobranch carbonic anhydrase is the archtypal vertebrate enzyme, as has been earlier suggested,² we have recently investigated the erythrocyte carbonic anhydrase from hagfish (*Myxine glutinosa*),⁴ belonging to the cyclostomes, which are considered to be the most primitive vertebrates. Since the molecular weight of the hagfish enzyme was determined to 29 000, we wanted to get further evidence for the idea that a carbonic anhydrase with a molecular weight of about 29 000 rather than a heavier one is the ancestral vertebrate enzyme. Therefore we have purified the erythrocyte carbonic anhydrase from lamprey (*Lampetra fluviatilis*), which also belongs to the cyclostomes.

Experimental. Blood was obtained from a vein of freshly caught lamprey by using a syringe with

heparin-treated needles. The erythrocytes were isolated by centrifugation (2000 *g*, 20 min, 5 °C) and washed by an equal volume of cold 0.9 % NaCl. Hemolysis was achieved by addition of a 5-fold volume excess of distilled water. The hemolysate was dialyzed over night against 0.01 M Tris–H₂SO₄, pH 7.0.

The purification was made on a *p*-aminobenzene-sulfonamide Sepharose column⁵ (2.0 × 20 cm, flow rate 15 ml/h, 5 °C) equilibrated with 0.1 M Tris–H₂SO₄, pH 7.0. Adsorbed hemoglobin to the gel was washed away by the above buffer containing 0.4 M Na₂SO₄ and elution was performed by 0.1 M Tris–H₂SO₄, pH 7.0 containing 0.6 M NaI. The carbonic anhydrase fractions were desalted on a Sephadex G-25 column (1 cm × 8 cm). The CO₂ hydration activity was measured according to Rickli *et al.*⁶ and the activity units are defined according to: Activity units = 10(*t*_b – *t*_c)/*t*_c, where *t*_b and *t*_c are the times for obtaining the color change of the indicator in uncatalyzed and catalyzed reactions, respectively. The homogeneity of the preparation was analysed by polyacrylamide gel electrophoresis⁷ (7.5 % acrylamide, 0.095 M Tris–glycine, pH 9.5). Isoelectric focusing was performed in an LKB Model 8101 focusing column⁸ in a pH gradient of 3.5 to 10. Molecular weight analysis was carried out by gel filtration on a Sephadex G-100 column (2.1 cm × 110 cm, 0.1 M KCl in 0.05 M Tris–HCl, pH 7.5). Using high molecular weight blue Dextran, the void volume, *V*_v, was determined to be 138 ml. The calibration proteins were eluted at the following volumes: Bovine serum albumin (*M*_w = 67 000⁹), 1.36 *V*_v; human carbonic anhydrase B (*M*_w = 28 900¹⁰), 1.88 *V*_v; and horse heart cytochrome *c* (*M*_w = 12 400¹¹), 2.44 *V*_v.

Results and discussion. A summary of the yield enzyme activity during the purification of lamprey carbonic anhydrase is shown in Table 1. From the affinity chromatography CO₂ hydration activity was only detected in a single peak. Analysis by polyacrylamide gel electrophoresis of the pooled fractions containing enzymic activity revealed only one band, suggesting high purity of the preparation. Isoelectric focusing of the hemolysate also

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Table 1. Purification of lamprey erythrocyte carbonic anhydrase. The data in this table are from one preparation from 13 ml of hemolysate.

Purification step	Total activity/ units × 10 ⁻⁴	Specific activity/ units/A _{280nm} × 10 ⁻³	Yield/%
1. Hemolysis	1.21	0.019	100
2. Dialysis	1.07	0.016	88
3. Affinity chromatography	0.38 ^a	23.5 ^a	31

^a After desalting of the enzyme.

showed only one active component, indicating that there is only one erythrocyte form of the lamprey carbonic anhydrase. The *pI* of the lamprey carbonic anhydrase was determined to be 5.2. After desalting of the enzyme fractions by gel filtration the enzyme was concentrated by lyophilization. By these procedures no inactivation of the enzyme was noted. If alternatively dialysis overnight was used to remove NaI, a 50 % loss of activity occurred. Concentration of the enzyme by vacuum dialysis also resulted in inactivation. In the purification of elasmobranch³ and hagfish⁴ carbonic anhydrases problems with inactivation of the enzyme during various dialysis steps were also noticed. Addition Zn(II) lead to reactivation of the inactivated hagfish carbonic anhydrase,⁴ but had no effect on the lamprey enzyme. Two gel filtrations of lamprey carbonic anhydrase resulted in elution after 1.87 and 1.88 void volumes, corresponding to the molecular weights of 29 200 and 28 900, respectively. This molecular weight is in agreement with that of the hagfish carbonic anhydrase which supports our earlier suggestion that the ancestral vertebrate carbonic anhydrase had a molecular weight of approximately 29 000.⁴

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